

Barrenazine A and B; Two New Cytotoxic Alkaloids from an Unidentified Tunicate

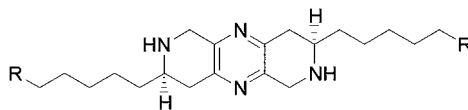
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ABSTRACT



1 Barrenazine A R= CH₂CH₃
2 Barrenazine B R= CH=CH₂

Two novel compounds, barrenazine A (1) and B (2), were isolated from an unidentified tunicate collected at Barren Islands, Madagascar. The two new compounds are of an unprecedented heterocyclic skeleton, namely 1,3,4,6,8,9-hexahydrodipyridino[3,4-*b*:3',4'-*e*]pyrazine. The structures of the two alkaloids were elucidated by interpretation of MS, COSY, HMQC, HMBC, NOESY, and ¹⁵N-HMBC data. Barrenazine A exhibits mild cytotoxicity against LOVO-DOX colon carcinoma (with a GI₅₀ value of 0.9 g/mL)

As part of our continuing program to discover bioactive compounds from marine invertebrates,^{1,2} we isolated two new cytotoxic alkaloids, having an unprecedented 1,3,4,6,8,9-hexahydrodipyridino[3,4-*b*:3',4'-*e*]pyrazine skeleton, designated as barrenazine A (1)³ and B (2).⁴ Barrenazine A is mildly cytotoxic to a few human cancer cells. In this paper we present the isolation and structure determination of these novel marine metabolites.

Pyrazine ring systems are not widely distributed in marine metabolites and are mostly restricted to a few distinct types

of structures. The most prevalent of these are the bis steroidal pyrazines comprised of the anticancer cephalostatins⁵ isolated from the African marine worm *Cephalodiscus gilchristi* and the cytotoxic ritterazines⁶ isolated from the tunicate *Ritterella tokioka*. Other occurrences of the pyrazine ring include the antibiotic compounds, pelagiomicins,⁷ isolated from the marine bacteria *Pelagibacte variabilis*, in which the pyrazine is part of a phenazine system; palythazin and isopalythazine which both consist of a hexahydrodipyranopyrazine,⁸ isolated from the zoanthid *Palythoa tuberculosa*; as well as clavulazine,⁹ a trihydropyranopyrazine metabolite isolated from the soft coral *Clavularia viridis*. Other examples of compounds containing a pyrazine nucleus include botrylla-

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(3) Barrenazine A (1): an oil; [α]_D²¹ –55.4 (c 0.48, MeOH); UV (MeOH) λ_{max} 288 (ε 6950), 313 (sh) nm; for ¹H and ¹³C NMR data, see Table 1; FABMS *m/z* 387 [M + H]⁺ (100), 287 (90); HRFABMS *m/z* 387.3476 (calcd for C₂₄H₁₃N₄, 387.3487).

(4) Barrenazine B (2): an oil; [α]_D²¹ –51.0 (c 0.26, MeOH); UV (MeOH) λ_{max} 288 (ε 6950), 313 (sh) nm; for ¹H and ¹³C NMR data, see Table 1; FABMS *m/z* 383 [M + H]⁺ (100), 285 (30); HRFABMS *m/z* 383.3103 (calcd for C₂₄H₁₃N₄, 383.3174).

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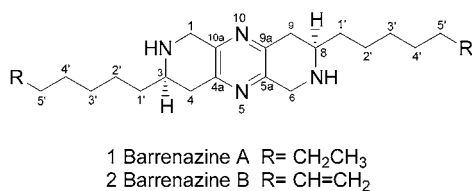
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zine A and B that were isolated from the red ascidian *Botryllus leachi*.¹⁰

The unidentified tunicate was collected at Barren Islands (Ban de l'Albatros), North-West of Madagascar.¹¹ The tunicate (40 g) was homogenized and extracted with chloroform–methanol (2:1) to give a brown gum (1.3 g). The extract was subjected to partition by the method of Kupchan et al.¹² The chloroform fraction was repeatedly chromatographed on Sephadex LH-20 columns, eluting with heptane–CHCl₃–MeOH (2:1:1) to afford **1** (15 mg, 0.037% dry weight) and **2** (3 mg, 0.0075% dry weight), as well as additional compounds with other side chains. Whether the real source of these new compounds is the sponge or symbiotic microorganisms is unknown.



The positive FAB mass spectrum of **1** exhibited a pseudomolecular ion [MH]⁺ at *m/z* 387. The molecular formula C₂₄H₄₂N₄ was determined by HRMS. The ¹³C NMR experiment revealed the presence of only 12 carbon atoms and thus it was deduced that **1** is of a symmetrical structure. A partial structure of barrenazine A could be constructed from HMQC, COSY, HMBC (Figure 1), and HSQC-TOCSY

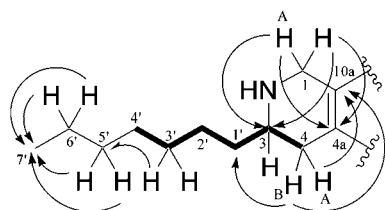


Figure 1. Key: HMBC (arrow with full arrowhead), COSY(—), and HSQC-TCSY (arrow with half arrowhead) correlations in barrenazine A.

data. A set of sequential COSY correlations was observed for protons H-4 through H-4'. HMBC correlations between C-7' and both H-6', H-5', as well as a correlation in the HSQC-TOCSY experiment between C-5' and H-3', allowed

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(11) The tunicate was collected at Barren Islands (Ban de l'Albatros) North-West of Madagascar (18°, 17', 260" south; 43°, 41', 200" east) on 5/2001 by scuba at a depth of 18 m. The tunicate is a massive colonial tunicate in orange balls. The colony of the tunicate is at rest with the state of the stolons. It is impossible to determine whether it is Polyclinidae or Polycitoridae. A voucher specimen is deposited at Museum d'Histoire Naturelle de Paris (voucher no. AM-737).

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the assignment of the C-4 to C-7' segment. ³J_{CH} couplings were observed between C-4a and both H-1A,1B, between C-10a and H-4A,4B, as well as between C-3 and H-1A,1B. Both C-3 and C-1 were positioned downfield (δ_C 44.8, 52.5 ppm) suggesting the presence of a tetrahydropyridino moiety.

This partial structure accounts for all the carbon and hydrogen resonances observed but only for one nitrogen atom. The UV spectrum of **1** showed an absorption at 288 nm (ε 6950) suggesting the presence of a pyrazine ring,¹³ which was in agreement with the two ¹³C NMR signals of C-4a and C-10a (144.9, 147.5 ppm).¹⁴ On the basis of this result and the symmetric nature of compound **1**, two possible symmetric structures (**I** and **II**) that satisfy the molecular formula could be suggested (Figure 2). The actual orientation

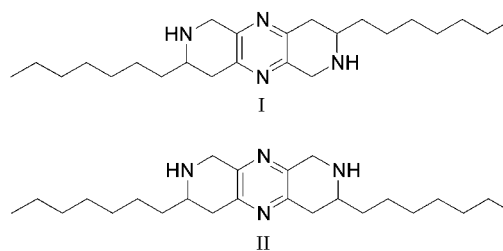


Figure 2. Two possible symmetric combinations that could apply for the structure of barrenazine A.

of the two 3-piperideines units with respect to the pyrazine ring could be determined on the basis of ¹⁵NH-HMBC experiment data. In the case of structure **I**, the ¹⁵NH-HMBC experiment was expected to give rise to NH correlations of two different nitrogen atoms, namely the nitrogen atoms of the pyrazine ring and the secondary amine moiety. In the case of structure **II**, however, the ¹⁵NH-HMBC experiment should give rise to NH correlations due to three different nitrogen atoms: two from the pyrazine ring and one from the amine moiety. ¹⁵NH-HMBC experiments were carried out with various delay times (60, 90, 120, and 150 ms). The best results were obtained when the delay was set to 120 ms and ³J_{NH} correlations were observed between H-1B(6B), H-4A,4B(9A,9B) and a single nitrogen atom resonating at 326 ppm (Figure 3) which is consistent with structure **I**. An

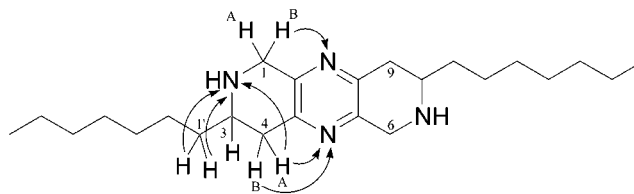


Figure 3. ¹⁵N-HMBC correlations in **1**.

additional set of important correlations observed in the ¹⁵NH-HMBC spectrum between H-4A,4B(9A,9B), H-1'A,1'B and

a nitrogen atom resonating at 49 ppm further support the structure suggested for barrenzine A.

Two diastereomers of **1**, both chemically and magnetically symmetric, depicted in Figure 4, agree with the above data

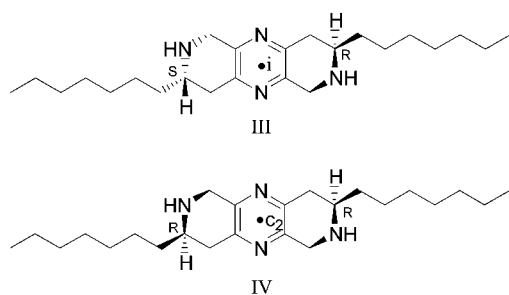


Figure 4. Two possible diastereomers of **1**.

and can be distinguished only by their optical properties. **III**, the S^*,R^* diastereomer, possesses a point of symmetry and thus should not display any optical activity while **IV**, the R^*,R^* diastereomer, exhibits C_2 symmetry and should display optical activity. Since **1** displayed optical activity $[\alpha]^{21}_D -55$ (and CD activity), it was established that the relative stereochemistry is R^*,R^* .

Generally, a half-chair is the predominant conformation for the piperidine ring.¹⁵ The signal of proton H-4B(9B) is a double-doublet presenting 18 Hz, 12 Hz couplings, due to coupling to H-4A and H-3 (and H-9B to H-9A and H-8). Therefore, both H-4B(9B) and H-3(8) must be quasi-axial and the two C-1'-C-7' heptyl chains must be quasi-equatorial. NOE correlations between H-3 and both H-4A and H-1A (and between H-3 and H-9A, H-6B) further corroborate this result. Moreover, Barrenzine A was converted to its tetramethyl derivative (**3**).¹⁶ From the ^1H NMR, ^{13}C NMR, and 2D experiments it became evident that two equivalent methyl groups were quasi-equatorial (δ_{H} 3.3 ppm, δ_{C} 52 ppm) and two other equivalent methyl groups were quasi-axial (δ_{H} 3.0 ppm, δ_{C} 44 ppm). The large upfield shift of the quasi-axial methyl groups resulted from γ effects by carbons C-10a(5a) and C-4(9). In a selective NOE experiment, a positive NOE enhancement was observed for the

quasi-equatorial methyl group upon irradiation of the H-3/8 protons, which were earlier determined to be in a quasi-axial position.

The positive mass spectrum of **2** exhibited a molecular ion $[\text{MH}]^+$ at m/z 383. The molecular formula, $\text{C}_{24}\text{H}_{38}\text{N}_4$, was determined by HRMS. The NMR data of **2** indicated it to be identical to **1** except for the presence of a terminal double bond (δ_{H} 5.32, 5.28, 5.76 ppm, δ_{C} 114.8, 138.7 ppm) in both chains of **2** while in **1** both chains are saturated.

Other similar compounds of the same heterocyclic skeleton but with different side chains are undoubtedly present in the tunicate. So far we were not able to separate mixtures of those compounds (some of which are isomers of barrenzine B with the double bond located at different sites of the heptyl chains).

Barrenzine A has been found to exhibit mild cytotoxicity against LOVO-DOX colon carcinoma with a GI_{50} value of 0.900 g/mL. A mixture of Barrenzine A and the above-mentioned similar compounds also exhibited mild cytotoxicity against LN-caP prostate carcinoma and K-562 Leukemia carcinoma with GI_{50} values of 0.594 and 0.180 g/mL, respectively. It is of note that the structurally related tetrahydropyridinopyrazine derivatives exhibit a variety of biological activities, i.e., inhibitory activity against various metalloproteinases¹⁷ and against keratinocyte growth¹⁸ and an inhibitory effect on TNF- α converting enzyme production¹⁹ and on matrix metalloproteinases (MMP).²⁰ These tetrahydropyridinopyrazines were also shown to be agonists of human melanocortin receptors²¹ and of interleukin 1 and therefore useful for the treatment of immunoinflammatory conditions.²² Therefore, synthesis of analogues with different side chains is in progress.

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Supporting Information Available: General experimental procedures and NMR data (^1H NMR, ^{13}C NMR, COSY, and HMBC) for barrenzines A and B. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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(16) Barrenzine A (5 mg) was dissolved in a mixture of acetone (0.2 mL) and methyl iodide (1 mL) and the mixture was heated to 70 °C for 96 h in a sealed reactive vial. After cooling, the mixture was freed from the solvent.